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Two new polyacetylenic compounds from *Atractylodes chinensis* (DC.) Koidz.

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Two new polyacetylenic compounds, (6E, 12Z)-tetradecadiene-8,10-diyne-1,3-diol (1) and (6Z, 12Z)-tetradecadiene-8,10-diyne-1,3-diol (2), were isolated from *Atractylodes chinensis* (DC.) Koidz. Their structures were established by analysis of spectroscopic data.

Keywords: Atractylodes chinensis (DC.) Koidz.; polyacetylenic compounds; (6E,12Z)-tetradecadiene-8,10-diyne-1,3-diol; (6Z,12Z)-tetradecadiene-8,10-diyne-1,3-diol

1. Introduction

The rhizome of Atractylodes chinensis (DC.) Koidz., an important Chinese traditional medicine, has been used to treat rheumatic diseases, digestive disorders, night blindness, and influenza [1]. Previous investigations showed that the polyacetylenic compounds were the characteristic and active constituents of A. chinensis, which had significant antiulcer activity toward some experimental gastric disorders [2], and strong antiinflammatory activity [3]. The present study on the phytochemistry of A. chinensis led to the isolation of two new polyacetylenic compounds, which were the Zisomers of (6E, 12E)-tetradecadiene-8,10diyne-1,3-diol [4] (Figure 1).

2. Results and discussion

Compound 1 was obtained as a pale yellow oil. The molecular formula was determined

to be $C_{14}H_{18}O_2$ by HR-TOF-MS at m/z 241.1204 $[M + Na]^+$. The ¹H NMR spectrum of **1** showed the presence of olefinic protons assignable to a *trans* and a *cis* double bond at δ 5.65 (1H, d, J = 16.2 Hz), 6.33 (1H, dt, J = 4.8, 16.2 Hz), and δ 5.56 (1H, d, J = 10.2 Hz), 6.17 (1H, m), respectively, and a methyl proton at δ 1.88 (3H, dd, J = 1.8, 7.2 Hz). ¹³C NMR spectrum showed 14 signals: one methyl, four methylenes, one methine, four olefinic carbons, and four acetylene carbons.

Analysis of the HMQC spectrum enabled us to allot the H-atoms to their bonded C-atoms (Table 1). The information concerning the location of these units was obtained from the HMBC experiment (Figure 2). The HMBC correlations from H-14 to C-12 and C-13, H-13 to C-11, and H-12 to C-10 suggested the presence of unit A (Figure 2). Another

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2 (6Z,12Z)-tetradecadiene-8,10-diyne-1,3-diol

Figure 1. Structures of compounds 1 and 2.

double bond H-7 correlated with C-5 and C-9, and H-6 correlated with C-8 and C-4, indicating the presence of unit B (Figure 2). The HMBC correlations from H-1 to C-2 and C-3, H-5 to C-3, and H-4 to C-2 established the unit C attached to C-4 (Figure 2). Finally, the acetylene carbon C-9 was connected to C-10. The structure could be further confirmed by COSY spectrum (Figure 3). Thus, compound **1** was characterized as (6E, 12Z)-tetradecadiene-8,10-diyne-1,3-diol.

Compound **2** was isolated as a pale yellow oil. The molecular formula was analyzed for $C_{14}H_{18}O_2$ by HR-TOF-MS at m/z 241.1197 [M + Na]⁺. The ¹H and ¹³C NMR spectral data of **2** indicated its structure to be closely related to that of **1**, with the difference in chemical shift and coupling constants of the four olefinic protons. The protons at δ 5.58 (1H, d,

J = 9.0 Hz, H-7) and 5.58 (1H, d, J = 9.0 Hz, H-12) indicated Z configuration for C-6-C-7 and C-12-C-13 double bonds. Thus, compound **2** was determined as (6Z,12Z)-tetradecadiene-8,10-diyne-1,3-diol.

3. Experimental

3.1 General experimental procedures

UV spectra were obtained on a Shimadzu UV-2201 spectrophotometer. IR spectra were recorded on an IFS-55 spectrophotometer. Optical rotations were determined on a Perkin-Elmer 241 MC polarimeter. The NMR spectra were run on a Bruker AV-600 spectrometer (600 MHz for ¹H and 150 MHz for ¹³C) in CD₃OD with TMS as an internal standard. The HR-TOF-MS data were obtained on a Waters LCT Premier XE time-of-flying mass

Table 1. 1 H and 13 C NMR spectral data of compound 1 in CD₃OD and key HMBC correlations (600 MHz for 1 H NMR and 150 MHz for 13 C NMR).

Position	$\delta_{ m C}$	$\delta_{ m H}$	HMBC
1	60.1	3.68 (2H, br s)	C-2, C-3
2	40.8	1.62 (1H, m)	C-1, C-3, C-4
		1.68 (1H, m)	C-1, C-3, C-4
3	69.1	3.72 (1H, br s)	C-2
4	37.5	1.55 (2H, m)	C-2, C-3, C-5, C-6
5	30.5	2.24 (1H, m)	C-3, C-4, C-6, C-7
		2.31 (1H, m)	C-3, C-4, C-6, C-7
6	149.3	6.33 (1H, dt, $J = 4.8$, 16.2 Hz)	C-4, C-5, C-8
7	109.9	5.65 (1H, d, $J = 16.2$ Hz)	C-5, C-6, C-9
8	81.7		
9	73.1		
10	79.1		
11	78.2		
12	110.0	5.56 (1H, d, $J = 10.2$ Hz)	C-10
13	143.4	6.17 (1H, m)	C-11, C-14
14	16.4	1.88 (3H, dd, $J = 1.8$, 7.2 Hz)	C-12, C-13

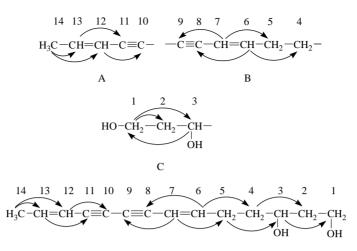


Figure 2. The key HMBC correlations of units A, B, C, and compound 1.

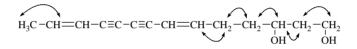


Figure 3. The key ${}^{1}H-{}^{1}H$ COSY correlations of compound 1.

spectrometer, and CD spectra on a Jasco CD-2095-plus spectrometer. Chromatography was performed on silica gel (200– 300 mesh; Qingdao Marine Chemical Group Co., Qingdao, China), Sephadex LH-20 (Pharmacia, Piscataway, NJ, USA), ODS (30–50 μ m; Tianjin Chemical Reagent Co., Tianjin, China), and preparative HPLC (Waters 600E pump, Waters 2489 UV spectrophotometric detector at 293 nm, Waters Sunfire Prep ODS reversed-phase column).

3.2 Plant material

The rhizomes of *A. chinensis* (DC.) Koidz., collected from Heilongjiang Province of China, were bought from the Cooperation of Traditional Chinese Medicine of Shenyang, China, in June 2006, and were identified by Prof. Jincai Lu (Shenyang Pharmaceutical University). A voucher specimen (20061005) has been deposited in the School of Traditional Chinese Medicine of Shenyang Pharmaceutical University, China.

3.3 Extraction and isolation

Dry rhizomes of A. chinensis (20.0 kg) were refluxed two times with 95% EtOH. The combined EtOH extracts were concentrated, suspended in H_2O , and then partitioned with petroleum ether (280.9 g), $CHCl_3$ (300.5 g), and EtOAc (42.0 g) successively to give three different polar parts. The CHCl₃ layer was subjected to silica gel column chromatography (CC) with a gradient of petroleum etheracetone (100:1-100:100). The fraction eluted with a solvent of petroleum etheracetone (100:10) (2.4 g) was further purified by Sephadex LH-20 CC eluted with $CHCl_3$ -MeOH (1:1) to yield a fraction (1.4 g), which was further subjected to ODS CC eluted with MeOH- H_2O . A fraction eluted with MeOH- H_2O (65:35) (22.1 mg) was further separated by preparative HPLC (60% MeOH, flow rate

Position	$\delta_{ m C}$	$\delta_{ m H}$	HMBC
1	60.2	3.69 (2H, br s)	C-2, C-3
2	40.7	1.62 (1H, m)	C-1, C-3, C-4
		1.68 (1H, m)	C-1, C-3, C-4
3	69.5	3.72 (1H, m)	C-2
4	37.7	1.55 (2H, m)	C-2, C-3, C-5, C-6
5	28.2	2.39 (1H, m)	C-3, C-4, C-6, C-7
		2.45 (1H, m)	C-3, C-4, C-6, C-7
6	148.7	6.16 (1H, m)	C-4, C-5, C-7, C-8
7	109.1	5.58 (1H, d, $J = 9.0 \mathrm{Hz}$)	C-5, C-6, C-9
8	79.7		
9	78.7		
10	79.7		
11	78.9		
12	110.0	5.58 (1H, d, $J = 9.0$ Hz)	C-10
13	143.5	6.16 (1H, m)	C-11, C-14
14	16.5	1.88 (3H, dd, $J = 1.8$, 7.2 Hz)	C-12, C-13

Table 2. ¹H and ¹³C NMR spectral data of compound **2** in CD₃OD and key HMBC correlations (600 MHz for ¹H NMR and 150 MHz for ¹³C NMR).

2.5 ml/min, wavelength 230 nm) to obtain compounds **1** (4.7 mg) and **2** (6.5 mg).

3.3.1 (6E,12Z)-Tetradecadiene-8,10diyne-1,3-diol (1)

Pale yellow oil, $[\alpha]_D^{20} - 24.0$ (c = 0.10, MeOH). IR (KBr) v_{max} (cm⁻¹): 3376, 2940, 1661, 1384, 1054, 1031; UV (MeOH) λ_{max} (nm): 229, 266, 276, 293, 312; CD (MeOH) $\Delta \varepsilon_{286.6 \text{ nm}} + 2.139$, $\Delta \varepsilon_{338.0 \text{ nm}} - 3.484$; ¹H and ¹³C NMR (CD₃OD) spectral data, see Table 1; HR-TOF-MS *m*/*z*: 241.1204 [M + Na]⁺ (calcd for C₁₄H₁₈O₂Na, 241.1204).

3.3.2 (6Z,12Z)-Tetradecadiene-8,10diyne-1,3-diol (2)

Pale yellow oil, $[\alpha]_D^{20} - 16.0 \ (c = 0.15, MeOH)$. IR (KBr) $v_{max} \ (cm^{-1})$: 3386, 2938, 1714, 1384, 1055, 1019; UV (MeOH) $\lambda_{max} \ (nm)$: 236, 266, 276, 293, 312; CD (MeOH) $\Delta \varepsilon_{286.6 \ nm} + 4.267$,

 $\Delta \varepsilon_{338.1 \text{ nm}} - 5.942$; ¹H and ¹³C NMR (CD₃OD) spectral data, see Table 2; HR-TOF-MS *m/z*: 241.1197 [M + Na]⁺ (calcd for C₁₄H₁₈O₂Na, 241.1204).

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